

# **CERTIFICATION**

# **AOAC®** Performance Tested<sup>SM</sup>

Certificate No.

020401

The AOAC Research Institute hereby certifies that the method known as:

# RapidChek® Listeria

manufactured by

Romer Labs 130 Sandy Drive Newark, DE 19713

This method has been evaluated in the AOAC® *Performance Tested Methods*<sup>SM</sup> Program and found to perform as stated by the manufacturer contingent to the comments contained in the manuscript. This certificate means that an AOAC® Certification Mark License Agreement has been executed which authorizes the manufacturer to display the AOAC *Performance Tested*<sup>SM</sup> certification mark along with the statement - "THIS METHOD'S PERFORMANCE WAS REVIEWED BY AOAC RESEARCH INSTITUTE AND WAS FOUND TO PERFORM TO THE MANUFACTURER'S SPECIFICATIONS" - on the above-mentioned method for a period of one calendar year from the date of this certificate (November 20, 2021 – December 31, 2022). Renewal may be granted at the end of one year under the rules stated in the licensing agreement.

November 20, 2021
Coates, Senior Director
Date

Scott Coates, Senior Director Signature for AOAC Research Institute

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SUBMITTING COMPANY Strategic Diagnostics Inc. 111 Pencaer Drive Newark, Delaware 19702 CURRENT SPONSOR Romer Labs Technology, Inc. 130 Sandy Drive Newark, Delaware 19713

KIT NAME(S)

RapidChek® Listeria

**CATALOG NUMBERS** 

Original catalog numbers: 3000019, 7000171, 7000174, 7000174P, 7000175, 7000175P, 7000175S, 7000176, 7000179, 7000180,7000182, 7000246

Updated catalog numbers: 10001173, 10001361, 10001364, 10001697, 10001365, 10001699, 10001700, 10001366, 10001369, 10001370, 10001372,

10001412

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APPLICABILITY OF METHOD

Target analyte - Listeria spp.

Matrixes – (25 g samples) - Ice cream, soft cheese, pasteurized whole milk, deli turkey, pepperoni, hot dogs, roast beef, cooked shrimp, smoked fish, potato salad, rubber (sponge,  $4 \times 4$  in), painted concrete (swab,  $1 \times 1$  in), stainless steel (swab,  $1 \times 1$  in)

Performance claims - The Lateral Flow Device (LFD) test Method for *Listeria* species was evaluated and was shown be equivalent to the reference methods.

REFERENCE METHODS

Hitchens, A.D. (1998) FDA, Bacteriological Analytical Manual; 8<sup>th</sup> Edition;

Chapter 10: Listeria monocytogenes. (2)

USDA/FSIS, (2006) Microbiology Laboratory Guidelines, Chapter 8; revision 5; Isolation and Identification of Listeria monocytogenes from Red Meat, Poultry, Egg, and Environmental Samples. (4)

## ORIGINAL CERTIFICATION DATE

June 15, 2004

CERTIFICATION RENEWAL RECORD

Renewed annually through December 2022.

### METHOD MODIFICATION RECORD

- 1. May 2008
- 2. December 2012 Level 1
- 3. May 2019 Level 1
- 4. November 2021 Level 1

SUMMARY OF MODIFICATION

- 1. Filter pad component change
- 2. Name change from Strategic Diagnostics to Romer Labs
- 3. Updated catalog numbers.
- 4. Updated USDA/FDA information.

Under this AOAC® Performance Tested  $^{\rm SM}$  License Number, 020401 this method is distributed by: NONE

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### PRINCIPLE OF THE METHOD (1)

The LFD Test for *Listeria* species can be used in combination with the proprietary RapidChek *Listeria* enrichment system for a rapid 40 hour test. After enrichment, an aliquot of sample broth is dispensed into a 12mm x 75mm test tube and boiled for 5 minutes. Once the sample has cooled, a test strip is added directly to the tube. The sample flows up the strip through a zone containing antibody coated colloidal gold reagents specific to *Listeria* species. If antigens are present in the sample, they will bind to the antibody-conjugates to form an antigen/antibody complex. As this complex migrates through the nitrocellulose matrix, it passes a zone of anti-*Listeria* antibody. If antigen is present, the complex is captured in this zone and is visualized by the formation of a red line. A second zone on the membrane is designed to capture any antibody-gold complex not bound in the first zone. As a result, when *Listeria* antigen is present, the formation of 2 red lines is observed, whereas when no *Listeria* is present, only 1 line forms.

#### **DISCUSSION OF THE VALIDATION STUDY (1)**

The RapidChek LFD and cultural methods have demonstrated excellent accuracy, sensitivity and specificity throughout these studies. Overall method agreement averaged 91% for the LFD and the cultural methods in food samples, though when method agreement data were adjusted to reflect the cases where the RapidChek methods recovered a greater number of positive samples than the reference methods, 100% and 102% method agreement were reported, respectively. In all 3 surface studies, the RapidChek methods reported greater numbers of positives than the reference method with the corresponding method agreement averaging 69% (131% when adjusted to reflect greater recovery). Overall, there were no false positive results and 2 false negative results observed using the LFD detection system (1% false negative rate, 0% false positive rate). There were 2 reported false positives and no false negative results using the RapidChek cultural method as the detection system with the food and surfaces tested (0% false negative rate, 3% false positive rate). In total, the RapidChek LFD and cultural methods reported 189 and 191 confirmed positive results, while the reference methods reported 169 confirmed results. The 2 methods have been shown to be capable of detecting very low levels (1cfu/25g) of *Listeria spp.* in a variety of food and has performed as well or better than the reference method with select environmental matrixes. Sample matrix effects on the LFD within the food and environmental samples examined in these studies were not apparent. Assay robustness studies indicate that the assay will perform under a wide range of environmental conditions. The assay is stable for at least a year at room temperature and results are highly reproducible from lot to lot.

	teria Isolates Source List – Inclu		1		
#	Species	Serotype	Strain	Source	LFD
		. 10			Activi
1	L. monocytogenes	1/2a	ATCC 51774 <sup>a</sup>	Human Blood	+
2	L. monocytogenes	1/2a	SDI 10-3b/c1 <sup>b</sup>		+
3	L. monocytogenes	1/2a	SDI 10a-3b/c2 <sup>b</sup>		+
4	L. monocytogenes	1/2a	SDI 11-3b/c3 <sup>b</sup>		+
5	L. monocytogenes	1/2a	USDA 472°		+
6	L. monocytogenes	1/2b	SDI 12-3b/c5 <sup>b</sup>		+
7	L. monocytogenes	1/2c	SDI 13-3b/c7 <sup>b</sup>		+
8	L. monocytogenes	1	ATCC 7644 <sup>a</sup>	Human	+
9	L. monocytogenes	2	ATCC 19112 <sup>a</sup>	Human Spinal Fluid	+
10	L. monocytogenes	3	ATCC 19113 <sup>a</sup>	Human	+
11	L. monocytogenes	3a	SDI 14-3b/c9 <sup>b</sup>		+
12	L. monocytogenes	3b	SDI 15-3b/d2 <sup>b</sup>		+
13	L. monocytogenes	Зс	SDI 16-3b/d4 <sup>b</sup>		+
14	L. monocytogenes	4a	SDI 17-3b/d6 <sup>b</sup>		+
15	L. monocytogenes	4ab	SDI 18-3b/d8 <sup>b</sup>		+
16	L. monocytogenes	4b	ATCC 13932 <sup>a</sup>	Human Spinal Fluid	+
17	L. monocytogenes	4b	ATCC 19115 <sup>a</sup>	Human	+
18	L. monocytogenes	4b	ATCC 43256°	Mexican Cheese	+
19	L. monocytogenes	4b	ATCC 51414 <sup>a</sup>	Raw Milk	+
20	L. monocytogenes	4b	SDI 19-3b/e1 <sup>b</sup>	ROW WIIK	+
21	L. monocytogenes	4b	U of G H7650 <sup>d</sup>		+
22	L. monocytogenes	4c	SDI 20-3b/e3 <sup>b</sup>		+
23	L. monocytogenes	40 4d	SDI 21-3b/e5 <sup>b</sup>		+
23 24	,	4u 4e	SDI 22-3b/e3		+
	L. monocytogenes	_			
25	L. monocytogenes	7	SDI 23-3b/e9 <sup>b</sup>	Oktober Leader	+
26	L. monocytogenes		SDI-51 <sup>b</sup>	Chicken Isolate	+
27	L. monocytogenes		U of G H7649 <sup>d</sup>		+
28	L. monocytogenes		SDI 201 <sup>b</sup>		+
29	L monocytogenes		SDI-52 <sup>b</sup>	Raw Beef	+
30	L. grayi		ATCC 19120 <sup>a</sup>	Chinchilla Feces	+
31	L. gray		ATCC 25401 <sup>a</sup>	Corn Stalks	+
32	L. innocua		USDA 15-666°		+
33	L. innocua	6a	ATCC 33090 <sup>a</sup>	Cow Brain	+
34	L. innocua	6b	ATCC 33091 <sup>a</sup>	Human Feces	+
35	L. innocua		SDI-3 <sup>b</sup>	Beef Isolate	+
36	L. innocua		SDI-198 <sup>b</sup>	Drain Sponge	+
37	L. innocua		SDI-53 <sup>b</sup>	Raw Turkey	+
38	L. seeligeri		ATCC 51334°	Clethrionomys glareolus Intestine	+
39	L. seeligeri	4a	ATCC 51335°		+
40	L. seeligeri		ATCC 35967 <sup>a</sup>	Soil	+
41	L. seeligeri		SDI 3BF1 <sup>b</sup>		+
42	L. seeligeri		SDI 3BF2 <sup>b</sup>		+
43	L. welshimeri	6b	ATCC 35897 <sup>a</sup>	Plant Material	+
<del>44</del>	L. welshimeri	6a	ATCC 43548 <sup>a</sup>		+
<del>45</del>	L. welshimeri	- Ou	SDI-199 <sup>b</sup>		+
46 46	L. welshimeri		SDI-50 <sup>b</sup>	Chicken Isolate	+
<del>40</del> 47	L. welshimeri		SDI-54 <sup>b</sup>	Smoked Salmon	+
47 48			SDI-54°	Jillokeu Jalilloli	+
	L. ivanovii				+
49	L. ivanovii	1	ATCC 700402 <sup>a</sup>		+

<sup>a</sup>American Type Culture Collection, Manassas, VA; <sup>b</sup>Strategic Diagnostics Inc. Culture Collection, Newark, DE; <sup>c</sup>Listeria Reference Laboratory, Donald S. Munro Collection;c; <sup>c</sup>United States Department of Agriculture, Wyndmoor, PA; <sup>d</sup>University of Georgia Culture Collection, Athens, GA.

Strain   BHI   RapidChek Listeria	Table 2	2: Non- <i>Listeria</i> isolates (1)		
2		Strain	ВНІ	RapidChek Listeria
3	1	Brochothrix thermosphacta 11509	-	-
A   Lactobacillus plantarum 8014   -   -	2	Citrobacter freundii 7A12	-	-
5         Micrococcus luteus 533         -         -           6         Rhodococcus equi ATCC 7698         -         -           7         Salmonella tyhpimurium 14028         -         -           8         Streptococcus mitis         -         -           9         Proteus vulgaris ATCC 6380         -         -           10         Bacillus cereus 11778         -         -           11         Enterococcus faecalis 19433         -         -           12         Staphylococcus aureus         +         -           13         Acinetobacter baumannii 19606         -         -           14         Aeromonas hydrophila #10         -         -           15         Chryseobacterium meningosepticum 13253         -         -           16         Klebsiella pneumoniae #9         -         -           17         Pseudomonas aeruginosa 10145         -         -           18         Myroides odoratus 4651         -         -           19         Vibrio spp. 62A1         -         -           20         Yersinia enterocolitica 23715         -         -           21         Lactobacillus coccus lacti 11454         -         -	3	Enterobacter cloacae #2	-	-
6 Rhodococcus equi ATCC 7698	4	Lactobacillus plantarum 8014	-	-
Salmonella tyhpimurium 14028   -   -	5	Micrococcus luteus 533	-	-
8         Streptococcus mitis         -         -           9         Proteus vulgaris ATCC 6380         -         -           10         Bacillus cereus 11778         -         -           11         Enterococcus faecalis 19433         -         -           12         Staphylococcus aureus         +         -           13         Acinetobacter baumannii 19606         -         -           14         Aeromonas hydrophila #10         -         -           15         Chryseobacterium meningosepticum 13253         -         -           16         Klebsiella pneumoniae #9         -         -           17         Pseudomonas aeruginosa 10145         -         -           18         Myroides odoratus 4651         -         -           19         Vibrio spp. 62A1         -         -           20         Yersinia enterocolitica 23715         -         -           21         Lactobacillus/coccus lacti 11454         -         -           22         Lactobacillus acidophilus 314         -         -           23         Pseudomonas spp.         -         -           24         E. coli 0157 35150         -         -	6	Rhodococcus equi ATCC 7698	-	-
9	7	Salmonella tyhpimurium 14028	-	-
10	8	Streptococcus mitis	-	-
11         Enterococcus faecalis 19433         -         -           12         Staphylococcus aureus         +         -           13         Acinetobacter baumannii 19606         -         -           14         Aeromonas hydrophila #10         -         -           15         Chryseobacterium meningosepticum 13253         -         -           16         Klebsiella pneumoniae #9         -         -           17         Pseudomonas aeruginosa 10145         -         -           18         Myroides odoratus 4651         -         -           19         Vibrio spp. 62A1         -         -           20         Yersinia enterocolitica 23715         -         -           21         Lactobacillus/coccus lacti 11454         -         -           22         Lactobacillus acidophilus 314         -         -           23         Pseudomonas spp.         -         -           24         E. coli 0157 35150         -         -           25         Hafnia alvei ATCC 25927         -         -           26         Enterococcus faecalis Biotype 43497         -         -           28         Corynebacterium diptheriae biotype 13         - <t< td=""><td>9</td><td>Proteus vulgaris ATCC 6380</td><td>-</td><td>-</td></t<>	9	Proteus vulgaris ATCC 6380	-	-
12   Staphylococcus aureus	10	Bacillus cereus 11778	-	-
12   Staphylococcus aureus	11		-	-
13         Acinetobacter baumannii 19606         -         -           14         Aeromonas hydrophila #10         -         -           15         Chryseobacterium meningosepticum 13253         -         -           16         Klebsiella pneumoniae #9         -         -           17         Pseudomonas aeruginosa 10145         -         -           18         Myroides odoratus 4651         -         -           19         Vibrio spp. 62A1         -         -           20         Yersinia enterocolitica 23715         -         -           21         Lactobacillus/coccus lacti 11454         -         -           22         Lactobacillus acidophilus 314         -         -           23         Pseudomonas spp.         -         -           24         E. coli 0157 35150         -         -           25         Hafnia alvei ATCC 25927         -         -           26         Enterococcus faecilis Biotype 6         -         -           27         Enterococcus faecilis Biotype 43497         -         -           28         Corynebacterium gibtheriae biotype 13         -         -           29         Enterococcus faecium biotype 1         - <td>12</td> <td>•</td> <td>+</td> <td>-</td>	12	•	+	-
15 Chryseobacterium meningosepticum 13253	13		-	-
15 Chryseobacterium meningosepticum 13253	14	Aeromonas hydrophila #10	-	-
16         Klebsiella pneumoniae #9         -         -           17         Pseudomonas aeruginosa 10145         -         -           18         Myroides odoratus 4651         -         -           19         Vibrio spp. 62A1         -         -           20         Yersinia enterocolitica 23715         -         -           21         Lactobacillus/coccus lacti 11454         -         -           22         Lactobacillus acidophilus 314         -         -           23         Pseudomonas spp.         -         -           24         E. coli 0157 35150         -         -           25         Hafnia alvei ATCC 25927         -         -           26         Enterococcus durans biotype 6         -         -           27         Enterococcus faecalis Biotype 43497         -         -           28         Corynebacterium diptheriae biotype 13         -         -           29         Enterococcus faecium biotype 11         -         -           30         Corynebacterium pseudogenitalium biotype         -         -           31         Enterococcus raffinosus biotype 5         -         -           32         Leuconostoc citreum biotype 109929	15		-	-
18	16		-	-
18  Myroides odoratus 4651	17	•	-	-
19 Vibrio spp. 62A1	18		-	-
20 Yersinia enterocolitica 23715	19	•	-	-
21 Lactobacillus/coccus lacti 11454	20		-	-
22 Lactobacillus acidophilus 314			-	-
23 Pseudomonas spp	22	·	-	-
24 E. coli 0157 35150	23	•	-	-
26 Enterococcus durans biotype 6	24		-	-
26 Enterococcus durans biotype 6	25	Hafnia alvei ATCC 25927	-	-
27 Enterococcus faecalis Biotype 43497	26		-	-
28 Corynebacterium diptheriae biotype 13			-	-
29 Enterococcus faecium biotype 11	28		-	-
Corynebacterium pseudogenitalium biotype  Intercoccus raffinosus biotype 5  Leuconostoc citreum biotype 109929  Streptococcus pyogenes 19615  Staphylococcus vitulinus biotype 265308  Bacillus subtilits biotype 987649  -	29		-	-
31 Enterococcus raffinosus biotype 5			-	-
32         Leuconostoc citreum biotype 109929         -         -           33         Streptococcus pyogenes 19615         -         -           34         Staphylococcus vitulinus biotype 265308         -         -           35         Bacillus subtilits biotype 987649         -         -		Enterococcus raffinosus biotype 5	-	<u>-</u>
33 Streptococcus pyogenes 19615  34 Staphylococcus vitulinus biotype 265308  35 Bacillus subtilits biotype 987649			-	<del>-</del>
34 Staphylococcus vitulinus biotype 265308			-	<u>-</u>
35 Bacillus subtilits biotype 987649		• • • • • • • • • • • • • • • • • • • •	-	-
			_	-
			-	

Table 4:Extramural	Comparison of the 4	0-h RapidChek <i>Listeria</i>	Methods with the	JSDA/FSIS and	FDA/BAM Referen	ce Methods: Roas	t Beef, Ricotta Ch	eese. (1)		
Sample Matrix	Method type	Strain	MPN /25g	# of Sample	Presumptive Positives	Confirmed Positives	Sensitivity (%)	Specificity (%)	Method Agreement (%)	Chi-Square
Roast Beef	RapidChek	Listeria innocua	0	5	0	0	n/a	100	100	n/a
	LFD 40 h		0.225 (0.05-0.93)* )*	20	14	14	100	100	75	1.45
	RapidChek		0	5	0	0	n/a	100	100	n/a
	Cultural 40 h		0.225 (0.05-0.93)* )*	20	14	14	100	100	75	1.45
	USDA/FSIS		0	5	0	0	n/a	n/a	n/a	n/a
	48 h		0.225 (0.05-0.93)*	20	9	9	n/a	n/a	n/a	n/a
Ricotta Cheese	RapidChek LFD 40 h	Listeria monocytogenes	0	5	0	0	n/a	100	100	n/a
		4ab	0.575 (0.17-2.0)*	20	12	12	100	100	95	0
	RapidChek Cultural		0	5	0	0	n/a	100	100	n/a
	40 h		0.575 (0.17-2.0)*	20	12	12	100	100	95	0
	FDA/BAM 48 h		0	5	0	0	n/a	n/a	n/a	n/a
			0.575 (0.17-2.0)*	20	13	13	n/a	n/a	n/a	n/a

Note: Roast Beef Aerobic Plate Count = 1.0 X 10^4 cfu/g Ricotta Cheese Aerobic Plate Count = <10 cfu/g

<sup>\* = 95%</sup> confidence Interval

Table 5: Extramu	ral Comparison of the	e 40-h RapidChek <i>Liste</i>	ria Methods with th	e USDA/FSIS	<b>Reference Method:</b>	Rubber environn	nental samples. (1	L)		
Sample Matrix	Method type	Strain	Nominal cfu /4 inch <sup>2</sup>	# of Sample	Presumptive Positives	Confirmed Positives	Sensitivity (%)	Specificity (%)	Method Agreement	Chi-Square
Rubber Surfaces	RapidChek LFD	Listeria monocytogenes	0	5	0	0	n/a	100	100	n/a
54.74555	40 h	4ab	1100	20	9	9	100	100	65	4.0
	RapidChek		0	5	0	0	n/a	100	100	n/a
	Cultural		1100	20	9	9	100	100	65	4.0
	USDA/FSIS		0	5	0	0	n/a	n/a	n/a	n/a
	48 h		1100	20	3	2	n/a	n/a	n/a	n/a

Sample Matrix	Method type	Strain	MPN /25g	# of Sample	Presumptive Positives	Confirmed Positives	Sensitivity (%)	Specificity (%)	Method Agreement	Chi-Square
Deli Turkey	RapidChek LFD	Listeria monocytogenes	0	5	0	0	n/a	100	100	n/a
	40 h	1/2a	0.4 (0.1-1.3)*	20	14	15	93	100	95	0
	RapidChek Cultural		0	5	0	0	n/a	100	100	n/a
	40 h		0.4 (0.1-1.3)*	20	15	15	100	100	90	0.1
	USDA/FSIS 48 h		0	5	0	0	n/a	n/a	n/a	n/a
			0.4 (0.1-1.3)*	20	13	13	n/a	n/a	n/a	n/a
Hotdogs	RapidChek LFD	Listeria welshimeri	0	5	0	0	n/a	100	100	n/a
	40 h		2.3 (0.6-9.5)*	20	16	16	100	100	100	0.13
	RapidChek Cultural		0	5	0	0	n/a	100	100	n/a
	40 h		2.3 (0.6-9.5)*	20	16	16	100	100	100	0.13
	USDA/FSIS 48 h		0	5	0	0	n/a	n/a	n/a	n/a
			2.3 (0.6-9.5)*	20	16	16	n/a	n/a	n/a	n/a

n/a = not applicable

Note: Deli Turkey Aerobic Plate Count = <9 X 10^3 cfu/g Hotdogs Aerobic Plate Count = < 8 10^3 cfu/g

<sup>\* = 95%</sup> confidence Interval

Table 7: Intramur	al Comparison of the	e 40-h RapidChek <i>Lister</i>	ia Methods with t	the USDA/FSI	S and FDA/BAM Ref	erence Methods	: Pepperoni and S	moked Fish. (1)		
Sample Matrix	Method type	Strain	MPN /25g	# of Sample	Presumptive Positives	Confirmed Positives	Sensitivity (%)	Specificity (%)	Method Agreement	Chi-Square
Pepperoni	RapidChek LFD	Listeria monocytogenes 3b	0	5	0	0	n/a	100	100	n/a
	40 h		1.1 (0.3-4.3)*	20	10	10	100	100	90	0.07
	RapidChek Cultural		0	5	0	0	n/a	100	100	n/a
	40 h		1.1 (0.3-4.3)*	20	10	10	100	100	90	0.07
	USDA/FSIS 48 h		0	5	0	0	n/a	n/a	n/a	n/a
			1.1 (0.3-4.3)*	20	12	12	n/a	n/a	n/a	n/a
Smoked Fish	RapidChek LFD 40 h	Listeria monocytogenes 7	0	5	0	0	n/a	100	100	n/a
	40 n	,	5.0 (1.5-17.8)*	20	14	14	100	100	90	0.1
	RapidChek Cultural 40 h		0	5	0	0	n/a	100	100	n/a
	40 11		5.0 (1.5-17.8)*	20	14	14	100	100	90	0.1
	FDA/BAM 48 h		0	5	0	0	n/a	n/a	n/a	n/a
			5.0 (1.5-17.8)*	20	16	16	n/a	n/a	n/a	n/a

Note: Pepperoni Aerobic Plate Count = 1.8 X 10^5 cfu/g
Smoked Fish Aerobic Plate Count = 1.89 X 10^5 cfu/g

<sup>\* = 95%</sup> confidence Interval

Table 8: Intramur	al Comparison of the	e 40-h RapidChek <i>Lister</i>	ia Methods with t	he FDA/BAM	Reference Method:	Cooked Shrimp,	and Potato Salad.	(1)		
Sample Matrix	Method type	Strain	MPN /25g	# of Sample	Presumptive Positives	Confirmed Positives	Sensitivity (%)	Specificity (%)	Method Agreement (%)	Chi-Square
Cooked Shrimp	RapidChek LFD	Listeria monocytogenes 3c	0	5	0	0	n/a	100	100	n/a
·	40 h	, ,	0.2 (0.1-0.9)*	20	11	11	100	100	100	0.1
	RapidChek Cultural		0	5	0	0	n/a	100	100	n/a
	40 h		0.2 (0.1-0.9)*	20	11	11	100	100	100	0.1
	FDA/BAM 48 h		0	5	0	0	n/a	n/a	n/a	n/a
			0.2 (0.1-0.9)*	20	11	11	n/a	n/a	n/a	n/a
Potato Salad	RapidChek LFD	Listeria monocytogenes	0	5	0	0	n/a	100	100	n/a
	40 h	1/2b	1.1 (0.2-4.3)*	20	16	17	94	100	90	0.17
	RapidChek Cultural		0	5	0	0	n/a	100	100	n/a
	40 h		1.1 (0.2-4.3)*	20	18	17	100	87	95	0
	FDA/BAM 48 h	]	0	5	0	0	n/a	n/a	n/a	n/a
			1.1 (0.2-4.3)*	20	18	18	n/a	n/a	n/a	n/a

Note: Cooked Shrimp Aerobic Plate Count = 1.8 X 10^4 cfu/g Potato Salad Aerobic Plate Count = 1.8 X 10^4 cfu/g

<sup>\* = 95%</sup> confidence Interval

Table 9: Intramu	ral Comparison of th	ne 40-h RapidChek <i>Liste</i>	ria Method with t	he FDA/BAM	Reference Method:	Whole Milk, and	Ice cream. (1)			
Sample Matrix	Method type	Strain	MPN /25g	# of Sample	Presumptive Positives	Confirmed Positives	Sensitivity (%)	Specificity (%)	Method Agreement (%)	Chi-Square
Whole Milk	RapidChek LFD	Listeria monocytogenes	0	5	0	0	n/a	100	100	n/a
	40 h	4b	5.3 (1.5-18.3)*	20	17	17	100	100	95	0
	RapidChek Cultural		0	5	0	0	n/a	100	100	n/a
	40 h		5.3 (1.5-18.3)*	20	17	17	100	100	95	0
	FDA/BAM 48 h	]	0	5	0	0	n/a	n/a	n/a	n/a
			5.3 (1.5-18.3)*	20	18	18	n/a	n/a	n/a	n/a
Ice Cream	RapidChek LFD	Listeria monocytogenes 3a	0	5	0	0	n/a	100	100	n/a
	40 h		5.8 (1.7-20.3)*	20	20	20	100	100	80	2.25
	RapidChek Cultural	]	0	5	0	0	n/a	100	100	n/a
	40 h		5.8 (1.7-20.3)*	20	20	20	100	100	80	2.25
	FDA/BAM 48 h	]	0	5	0	0	n/a	n/a	n/a	n/a
			5.8 (1.7-20.3)*	20	16	16	n/a	n/a	n/a	n/a

Note: Whole Milk Aerobic Plate Count = 9 X 10^3 cfu/g Ice Cream Aerobic Plate Count = < 9 X 10^3 cfu/g

<sup>\* = 95%</sup> confidence Interval

Table 10: Intram	ural Comparison of t	he 40-h RapidChek <i>List</i>	eria Methods with	the USDA/FS	IS Reference Metho	d: Painted Concr	ete, and Stainless	Steel. (1)		
Sample Matrix	Method type	Strain	Nominal MPN /inch²	# of Sample	Presumptive Positives	Confirmed Positives	Sensitivity (%)	Specificity (%)	Method Agreement (%)	Chi-Square
Painted Concrete	RapidChek LFD	Listeria monocytogenes 4b	0	5	0	0	n/a	100	100	n/a
	40 h		1X10 <sup>4</sup>	20	18	18	100	100	60	6.13
	RapidChek Cultural	1	0	5	0	0	n/a	100	100	n/a
	40 h		1X10 <sup>4</sup>	20	19	18	100	86	60	6.13
	USDA/FSIS 48 h	1	0	5	0	0	n/a	n/a	n/a	n/a
			1X10 <sup>4</sup>	20	10	10	n/a	n/a	n/a	n/a
Stainless Steel	RapidChek LFD	Listeria innocua	0	5	0	0	n/a	100	100	n/a
	40 h		5.5X10 <sup>2</sup>	20	18	18	100	100	82	0.57
	RapidChek Cultural 40 h		0	5	0	0	n/a	100	100	n/a
	40 11		5.5X10 <sup>2</sup>	20	18	18	100	100	82	0.57
	USDA/FSIS 48 h		0	5	0	0	n/a	n/a	n/a	n/a
			5.5X10 <sup>2</sup>	20	15	15	n/a	n/a	n/a	n/a

#### **DISCUSSION OF MODIFICATION APPROVED MAY 2008 (3)**

The RapidChek method has demonstrated excellent sensitivity, specificity, and accuracy in the detection of *Listeria* species in food and environmental surfaces throughout these studies. The RapidChek method reported greater numbers of positive results than the reference method in all three matrixes. Overall, there were no false positives results and two false negative results observed using the lateral flow device. In total, the RapidChek method reported 35 confirmed positive results, while the reference method reported 26 confirmed positive results. Both methods have been shown to be capable of detecting very low levels (0.575 cfu/25g) of *Listeria* spp. in food and the RapidChek method has performed as well or better than the reference method with the selected environmental surfaces.

ble 1. Re	sults from	the Hot Dog I	Method Con	nparison (3)								
Matrix	Analyte	Method	Number of Samples	Inoculation Level, MPN/25g	Presumptive Positives	Confirmed Positives	Reference Method	Chi- Square	Sensitivity Rate	False Negative Rate	Specificity Rate	False Positive Rate
Hot Dogs	L. mono 4d	RapidChek	5	0	0	0	0	-	-	-	100	0
	M21		20	0.575	11	12	11	0	92	8	-	
ble 2. Re	sults from	Plastic Surfac	e Method C	omparison (3)								
Matrix	Analyte	Method	Number of Samples	Nominal cfu/4inch²	Presumptive Positives	Confirmed Positives	Reference Method	Chi- Square	Sensitivity Rate	False Negative Rate	Specificity Rate	False Positive Rate
Plastic	L. innocua ATCC	RapidChek	5	0	0	0	0	-	-	-	100	0
	33090		20	1.00E+03	13	14	9	1.58	93	7	-	-
ble 3. Re	sults from	Stainless Stee	l Method C	omparison (3)								
Matrix	Analyte	Method	Number of Samples	Nominal cfu/4inch <sup>2</sup>	Presumptive Positives	Confirmed Positives	Reference Method	Chi- Square	Sensitivity Rate	False Negative Rate	Specificity Rate	False Positive Rate
Steel	L. mono 4b ATCC	RapidChek	5	0	0	0	0	-	-	-	100	0
	13932		20	3.00E+02	9	9	6	0.94	100	0	-	-

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